

Immobilization of biomolecules on natural clay minerals for medical applications

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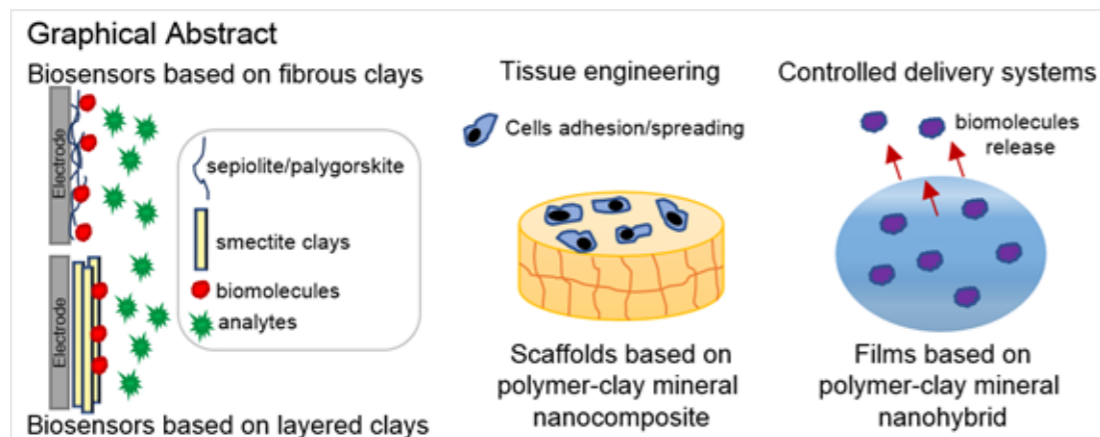
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ABSTRACT

Biomolecules are a group of organic entities that are important in many areas of research on nanomaterials and for biomedical and pharmaceutical applications. Advanced systems have been developed to attempt to protect the activity of biomolecules from rapid degradation and instability. Among these techniques, the incorporation or immobilization of biomolecules has become popular in the development of biocomposites. As such, clay minerals appear to be promising materials; combining a nanometer-scale size with their adsorptive capacity, lack of toxicity, and biocompatibility would result in enhanced biomaterial properties. This mini-review discusses the recent advances concerning biological molecules immobilized on clay minerals and their biomedical applications as biosensors, in regenerative medicine, and even as controlled delivery systems.



1. Introduction

Biomolecules are a group of organic entities of biological origin, such as polysaccharides, lipids, vitamins, enzymes, amino acids, peptides, proteins, and nucleic acids. A variety of materials have been proposed with properties derived from the functions of these molecules.^{1,2} However, systems containing biomolecules often show restricted recovery and reuse because of their lack of stability at elevated temperatures, in organic solvents, and in a gastrointes-

tinal environment.³

Biocomposites are materials based on an inorganic solids, such as clay minerals, in association with organic compounds.⁴ This approach can protect biomolecules from degradation in arrays derived from natural resources.⁵ In addition, molecules immobilized on nanosystems show well-preserved catalytic activity and enhanced properties.^{6,7} Systems containing biomolecules have a wide variety of applications in clinical or industrial use; thus,

inorganic solids and their assemblies play an important role as immobilizing matrices.⁸

Clay minerals are solids of the phyllosilicate family with the potential for biomolecule immobilization, resulting from their specific physicochemical characteristics, such as high surface reactivity.⁴ In addition, these materials exhibit antimicrobial properties with good biocompatibility.^{9,10} The advantages of these materials include their abundance, low cost, and potential as regional products.⁴

According to Jayrajshin and coworkers¹¹, the interaction between nanoclays and organic compounds has been studied in different areas of research, such as engineered nanomaterials for biomedical and pharmaceutical applications. The proposed applications for nanocomposites include the use of biomaterials as scaffolds, drug carriers for delivery systems, patches for wound healing, and as modifiers of biological substrates in electronic or implantable devices.

The present review focuses on studies related to different types of biological molecules immobilized on natural clay minerals published in the last ten years. We review the main experimental research in the biomedical application of clay nanocomposites, including their use as biosensors and controlled delivery systems and in regenerative medicine.

2. Clay minerals – major characteristics

Aluminosilicates, such as montmorillonite, kaolinite, illite, sepiolite, and palygorskite (or attapulgite) belong to two types of abundant inorganic solids in nature.¹² There are certain differences between layered and fibrous clay minerals, i.e., expanding and non-expanding types. As an example of layered clay, montmorillonite shows expansibility, 1-nanodimensional particle, high charge density and cation exchange capacity, low-density silanol groups at the edges, and particles in layered stacks. By contrast, fibrous clays, such as sepiolite and palygorskite, exhibit a non-swelling process, 2-nanodimensional particles, low charge density and cation exchange capacity, high-density silanol groups ($\equiv\text{Si-OH}$) at the broad external surface area, and particles in bundles.¹³

Both classes are micro and nano-sized¹⁴, and present strong adsorption strength and ion exchange abilities, and a high internal surface area. The layers or sheets are constituted by basic arrangements of clay mineral tetrahedral silicates and octahedral hydroxide sheets, giving rise to various classes of clay minerals of type 2:1 or 1:1.¹⁰ The permanent net negative charge of the layers or sheets, resulting from the substitution of Al^{3+} by Fe^{3+} or Mg^{2+} , the presence of hydroxyls at the limits, and compensatory cationic ions located in interlayer/sheets spaces are responsible for cation exchange abilities. Broken edges of clay show pH-dependence, staying positive at low pH and negative at high pH, originating from the surface reactivity of the clay mineral.^{14, 10}

Thus, biomolecules, either negatively or positively

charged, can be immobilized on the surface, edges, or interlayer/microchannels of clay particles.^{10,15} Adsorption of organic molecules on clay minerals reported in the literature¹⁶ includes hydrophobic interactions, hydrogen bonding, protonation, ligand exchange, cation exchange, cation bridging, and water bridging.

Natural clay minerals to be used for medical purposes must be purified to eliminate impurities, such amorphous or organic materials.⁶ Preparation of the purified clay sample is beneficial, because the final product is of very high purity. Despite the costs incurred during the purification of natural clay minerals, which are used for medical applications, they remain an attractive choice.

3. Diversity of biomolecules associated with clay minerals

Many biological agents incorporated to clay minerals can be released in organic systems for a range of biomedical applications. The diversity of biomolecules used to form new materials in association with natural clay minerals, as well as the forms of incorporation and the state of the molecules, are summarized in table 1.

As seen in table 1, clay minerals can adsorb various biomolecules, including proteins, nucleic acids, and carbohydrates. The majority of the reports on the use of biomolecules refer to polymer and enzyme immobilization. In the field of release systems, the number of reports on the use of clay minerals has increased in the last four years. Controlled drug delivery systems allow temporal and/or spatial control of release rates, thus, allowing acceleration, delay, or prolongation of release, as well as site-specific targeting.⁴

Among different types of clay minerals, montmorillonite has been studied more frequently in biological applications, possibly because of its high ion exchange capability and because it is widely distributed in nature. Chen and coworkers⁵⁰ reported that effective intercalation of proteins within the galleries of montmorillonites can be achieved via space enlargement and exchange processes, while retaining the native conformation of the guest proteins and the multilayered structure of the bioinert host plates. Despite this, according to Ruiz-Hitzky and coworkers,⁴ in certain cases, fibrous clays are more interesting than layered clays and can display higher enrichment of the mechanical properties of biomaterials.

Furthermore, bionanocomposites can be processed as films or as porous cellular materials, using solvent casting and freeze-drying processes. The following sections discuss the biomedical applications of clay minerals.

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Table 1 - Diversity of immobilized biomolecules on natural clay minerals reported in the literature in the last ten years.

Biomolecule	Clay/clay mineral	Form of Incorporation	Condition of the molecule in the clay	Ref.
Carrageenan	montmorillonite	Suspension in solution	Intercalated	17
		Electrospinning from mixing in solution	Intercalated by ion exchange	18
Chitosan		Mixing in solution	Intercalation with a planar conformation	19
		Dispersion in acetic acid aqueous solution	Intercalated by ion exchange	20
Chitosan-PVA		Electrospinning for mixing in solution	Intercalated by ion exchange	21
Alginate/Ghatti Gum		Dispersion in aqueous solution	Intercalated complexes	22
Vitamin B12		Adsorption in phosphate-buffered saline (PBS) buffer solution	Intercalated with diffusion of vitamin B12 to the interlayer spaces	23
Cellulose		Solution mixing process	Intercalation	24
Lipid		Aqueous solution of clays added in an emulsion containing the lipid to form microparticles	Not determined; however, clay exfoliation was proposed to form the microparticles, as observed by scanning electron microscopy (SEM).	25
Pectin/chitosan		montmorillonite with silica gel and sepiolite	Solubilization of chitosan, followed by addition of the clay under ultrasound, and formation of the hydrogel with the addition of pectin under vigorous shaking	Amorphous phases were identified.
Carboxymethyl-starch (CMS)	Clay was dispersed in water and stirred with CMS, glycerol, and citric acid. The mixture was poured into a polytetrafluoroethylene (PTFE) mold to form a film		Intercalated	27
Cysteine, aspartic, glutamic acids	Suspension in solution		Intercalated	28
Dendrimeric peptide	Suspension in solution		Intercalated	29
Cysteine, thiourea, and thiocyanate	Suspension in seawater		Adsorption with a smaller expansion of the layer	30
DNA/RNA	Adsorption in double-deionized water followed by precipitation in a microcentrifuge		Intercalated	31
Poly lactide	Melt extruded		Dispersed in polymer	32
Proteins	Adsorption on buffer solution		Not indicated in the text	33
Hemoglobin or methyl viologen	Clay films were prepared by dropping a known volume of clay colloids – soaked in methyl viologen, or by dropping of Hb/ clay aqueous mixture onto the electrode		Not indicated in the text	34

Table 1 - Diversity of immobilized biomolecules on natural clay minerals reported in the literature in the last ten years(cont.)

Hemoglobin (Hb) or methyl viologen	saponite	Clay films were prepared by dropping a known volume of clay colloids – soaked in methyl viologen, or by dropping of Hb/ clay aqueous mixture onto the electrode	Not indicated in the text	34
L-DOPA (precursor of dopamine)		Adsorption in aqueous solution at pH 7.5	Intercalated – theoretical and experimental study	35
Chitosan		Films were prepared from the solubilization of the chitosan and saponite dispersion in the presence of glycerol	Coplanar alignment of saponite nanoplatelets with two monolayers of chitosan macromolecules in the gap	36
Cysteine, thiourea, thiocyanate	bentonite*	Suspension in seawater	Adsorption with a smaller expansion of the layer	30
Amino acids		Suspension in seawater	Intercalated	37
<i>Homalomena aromatica</i> rhizome oil		Clay was modified with oil, followed by modification with epoxy resin, and cured	Intercalated	38
Amino acids	kaolinite	Suspension in seawater	Intercalated	37
Gellan gum	halloysite	Hydrogel was prepared with dispersion of gellan gum, followed by clay addition under sonication	Not indicated in the text	39
Pectin, cellulose, chitosan		Adsorption	Not indicated in the text	40
Cellulose		Halloysite nanotubes were incorporated into a cellulose NaOH/urea solution to prepare composite hydrogels by epichlorohydrine crosslinking	Not indicated in the text	41
Carrageenan		Film was prepared from a mixture of aqueous solution of Carrageenan and of clay, followed by the addition of glycerol	Carrageenan interacted with the hydroxyl surface groups of clay	42
Starch, alginate	sepiolite	Polymers were dissolved and added to a dispersion of clay, and then mixed using a processor	Strong interactions were indicated by the considerable perturbation of the stretching vibration band of –OH in the silanol groups	43
Chitosan		Chitosan was dissolved in acetic acid, clay was dispersed in water, and the solutions were mixed to form a film	The materials interact on the surface of the clay without penetration inside the tunnels	44
Starch and alginate or chitosan		Dispersion of clay in the solution containing the polymer	Not indicated in the text	15
<i>Arabinoxylan</i>		Polymer and clay were solubilized and suspended in water separately, and mixtures were degassed by ultrasonication and then added onto plates to form a film	The SEM images showed a good dispersion of clay in the films	45
Xanthan gum		Polymer and clay were solubilized and suspended in water separately, and mixtures were stirred to obtain a gel and lyophilized for solvent removal	Polymers interact on the surface of clay via hydrogen-bonding interactions	46

Table 1 - Diversity of immobilized biomolecules on natural clay minerals reported in the literature in the last ten years(cont.)

Poly(lactic-co-glycolic acid) - PLGA	palygorskite (attapulgitite)	PLGA was dissolved in a mixed solvent of tetrahydrofuran– dimethylformamide, clay was added to form a homogeneous solution for electrospinning	Not indicated in the text	47
Protein zein	palygorskite with sepiolite	Adsorption in ethanol/water media	External surface	48
Sodium alginate	palygorskite (attapulgitite)	Alginate was dissolved in water; for crosslink formation, ammonium persulfate (initiator) was added. Acrylic acid was neutralized with NaOH, and completely mixed with crosslinker, N,N'-methylene-bis-acrylamide, for incorporation of different amounts of clay into the hydrogel formed, and the samples were dried to obtain the nanocomposites	Not indicated in the text	49
Chitosan and sodium alginate	palygorskite (attapulgitite)	Hydrogel was prepared with different concentrations of palygorskite by graft-copolymerization in association with acrylic acid. After alginate addition, the mixture was dripped in calcium chloride solution to obtain crosslinked microparticles in Ca ²⁺	Not indicated in the text	50

*The term “bentonite” refers to a mixture of minerals from the smectites group with a predominance of montmorillonite clay mineral; however, we prefer to use the term as reported in the articles cited in the table.

4. Applications of biomolecules immobilized on clay minerals

4.1. Biosensors

Biosensors are chemical sensors in which the recognition system uses a biological mechanism to measure the interaction between the analyte and the sensor device, transforming quantitative or qualitative information into a measurable electrical signal.⁵¹

Clay minerals have been used as supports or modifiers of substrates in the field of electronic devices for electroanalytical purposes. These inorganic solids have received increasing attention since they were first used in these systems by Ghosh and Bard in 1983⁵², who reported the first electrode modified with a clay film. Following the use of natural zeolite on modified carbon paste electrodes for analytical purposes, the clay mineral sepiolite has also been applied for the same purpose⁵³.

Some studies have described the use of clays as electrode modifiers or as clay-containing matrices.⁵⁴⁻⁵⁸ However, only a few have studied the biomedical applications of clay biosensors or the diversity of biomolecules associated with clay minerals.

These materials can be used in electroanalysis because of their electrocatalytic properties and capacity to immobilize biocatalytic entities to improve the sensitivity and/or selectivity of the detection process.^{3,53,59} Biomolecules immobilized on clay minerals supports to develop biosensors that have been reported in literature are

summarized in table 2.

Nanostructured materials can be formed by the intercalation of organic molecules, such antibodies, peptides, proteins, genes, bacteria, cells, polymers, or enzymes within the interlayer, edges, or surface space of the clay minerals.⁵⁶ Notably, reports show that enzymes are the most frequently immobilized entities on clay matrices, including their use as amperometric biosensors, because of the sensitivity and specificity of their chemical reactions in these systems. For example, enzymatic biosensors for the detection of catechol, glucose, hydrogen peroxide, and phenol have been developed.

Immobilization should ensure that the biological activity of the immobilized biomolecule is maintained and its stability is preserved or enhanced while providing accessibility to the analytes. In this regard, clay minerals have proved to be suitable materials.¹⁰

Palygorskite represents an excellent inorganic material for the development of biosensors because of its electrocatalytic activity, which may be attributed to its high adsorption capability and the presence of OH groups on its surface. These features allow electron transfer between the electrode and the detected analytes.⁷⁵ Furthermore, its large surface area, high biocompatibility, and stability make it a promising material for enzyme immobilization.⁶⁸

Recently, halloysite nanotubes have been developed by evaporative assembly. They are promising natural materials because of their rough surfaces, which provide higher cancer cell capture efficiency compared with blank

Table 2 - Biosensors based on clay modifier electrodes and immobilized biomolecules.

Biomolecules	Clay modifiers	Electrode	Biosensor	Ref
Pyranose oxidase	montmorillonite/polyglycolide (PGA)	glassy carbon	glucose	60
Cisteyne	bentonite-AuNanoparticles	glassy carbon	ascorbic, uric and folic acid	61
Enzymes	halloysite	glass capillary	cancer cells	62
Horseradish peroxidase	sepiolite/carbon nanotubes (CNT)/PVA	-	peroxidase	63
Glucose oxidase	montmorillonite/ Gly, Lys, Glu	glassy carbon	-	64
Bovine serum albumin (BSA), glutaraldehyde (GA) and pyranose oxidase	montmorillonite/ calixaren-NH ₂	glassy carbon	-	65
Laccase	montmorillonite/ histidine	glassy carbon	phenol	66
Glucose oxidase	palygorskite-poly(o-phenylenediamine)/glutaraldehyde	-	glucose	67
Horseradish peroxidase	palygorskite	glassy carbon	hydrogen peroxide	68
<i>Lactobacillus bulgaricus</i> , <i>Streptococcus thermophilus</i>	palygorskite	oxygen	lactate	69
Horseradish peroxidase	palygorskite	glassy carbon	hydrogen peroxide (cellular reactive oxygen species)	70
Tyrosinase	palygorskite	glassy carbon	phenol	71
Hemoglobin	nontronite, montmorillonite and saponite/Fe ₂ O ₃	-	hydrogen peroxide	33
Glucose oxidase	palygorskite	glassy carbon	glucose (blood and urine samples)	72
Hemoglobin	palygorskite	glassy carbon	hydrogen peroxide	73
Cytochrome c	palygorskite	glassy carbon	hydrogen peroxide	74

capillary glass surfaces.⁶³ Their tubular structure make them suitable candidates for biomolecule capture and development of enzymatic biosensors.

The challenges regarding the development of biosensors based on nanocomposites include the ability of detecting lower concentrations of the analyte of interest, often at the trace level, to ensure the selectivity, sensitivity, and reproducibility of the system.⁵⁹

4.2. Regenerative medicine

Different strategies are required to develop a biomaterial, such as a suitable scaffold, which satisfies the requirements of cells in a three-dimensional support system or as a delivery vehicle incorporating bioactive compounds.⁷⁷

Hydrogels, containing including natural polymers, such as chitosan, gelatin, starch, and recently gellan

gum,⁷⁸ act as integrated networks of scaffolds because of the structural similarity of these components and have the potential to regulate cellular responses. However, their use has some limitations, such as relatively poor mechanical properties, high water sensitivity, or limited ability to support cell adhesion.^{4,7} These difficulties can be overcome by modification of their structure or the incorporation of bioactive molecules, such as proteins, peptides, or clay minerals.⁷

Polymer-clay mineral nanocomposites can contribute to this field because of their high porosity and compressive strength, which remains an ongoing challenge in scaffold design, particularly in bone repair.¹⁴ Another challenge is retaining the growth factors in the matrix in the gel network.⁷ In this regard, clay concentrations under 5% (w/w) have shown improvements in the modulus and strength of 3-D materials.¹⁴

Reviews concerning experimental clay research in regenerative medicine were carried out by Dawson and Oreffo¹⁴ Ruiz-Hitzky¹³, Chrzanowski⁷, and Bramhill and coworkers.⁷⁷ In the last four years, about 140 studies have been published concerning the use of clays in these systems, demonstrating the growing interest in this area. Most of the reports concerned montmorillonite and halloysite; however, other used kaolinite, palygorskite, and sepiolite.

Researchers have also examined the cellular response to biomaterials. Among them, Barua and coworkers³⁸ developed a polymeric matrix based on *Homalomena aromatica* rhizome oil-modified bentonite, which possessed antimicrobial activity. Biocompatibility assays were performed after subcutaneous implantation in Wistar rats. The biomaterial stimulated the adhesion and proliferation of dermatocytes, without any signs of toxicity.

Another study by Mohd and coworkers,⁷⁸ described the use of sodium montmorillonite (Na-MMT) modified with trimethyl ammonium bromide (CTAB-MMT) incorporated into a gellan gum (GG) hydrogel to improve its thermal stability. Cell studies showed that the Na-MMT composite was non-cytotoxic to skin fibroblast cells (CRL2522). In contrast, hydrogels with CTAB-MMT caused death and growth depletion of cells after 72 h.

Another advantage of fibrous clays compared with layered silicates is their very high density of silanol groups, which allows hydrogen bonding in addition to Van der Waals forces at the polymer-silicate interface.¹³ The incorporation of palygorskite nanorods into poly (lactic-co-glycolic acid) matrices contributed to the osteogenic differentiation of cells, without changing the uniform morphology and hemocompatibility of the scaffolds.⁴⁸

Another study by He and coworkers,⁷⁹ showed the use of natural nanopalygorskite to enhance vero cell productivity, without inducing cytotoxicity. This result suggested a useful strategy to reduce the cost of producing mammalian cell cultures for large-scale tissue engineering.

According to Bramhill and coworkers,⁷⁷ classical research has focused on bone regeneration; however, recent advances have also enabled the use of clay minerals at the soft tissue sites in the body. For these purposes, greater control of the physico-chemical properties of the biomaterials and their interactions at the body sites need to be evaluated. Future studies might focus on electro-spinning techniques or deposition in layers to develop new nanocomposite materials.⁷⁷

4.3. Controlled Release Systems

Biofunctional molecules, such as cells, nucleic acids, proteins, or lipids can be successfully stabilized while maintaining their biofunctional properties by being preserved in controlled release systems that are stabilized using clay minerals.

Ruiz-Hitzky and coworkers⁴ and Zafar and coworkers⁸⁰ reviewed biopolymer-clay nanocomposites for their

application as drug or biomolecule delivery systems, which rely on their properties of bioadhesion, biodegradability, and cell uptake. These features contribute to maintaining a constant dosage of the bioactive substance within the therapeutic dosage throughout the treatment course.

An experimental study reported the use of nanostructured montmorillonite clay, for controlling lipase-mediated digestion of medium chain triglycerides, engineered by spray drying oil-in-water emulsions.²⁵ The performance of montmorillonite-lipid microparticles (75% w/w) under simulated intestinal conditions suggested their use a novel biomaterial and that encapsulation optimized lipase efficiency as a smart delivery system for lipophilic biomolecules.

In recent studies, a sustained release system, based on chitosan and montmorillonite prepared by ion exchange, for controlled oral mucosal administration of chlorhexidine (CLX) was proposed. *In vitro* release tests showed sustained long-term release without an initial burst release.¹⁹

Another approach involves the use of bioactive film-forming matrices with improved functionality as wound dressings; these matrices allow controlled release of biomolecules.⁴⁰ Bionanocomposites based on polysaccharide-clay minerals can be used to encapsulate biomolecules lacking cytotoxicity, increasing mucoadhesivity and stimulating cell proliferation.⁸⁰

For bioactive film matrices, most studies have used lamellar silicates; however, recent studies confirmed that fibrous clay minerals are also promising for the development of bionanocomposites.^{4,81} A challenge in films manufacture is the fabrication of the nanocomposite itself.⁸⁰ Improvement of the characteristics of the new materials based on fibrous clay minerals can be obtained by suitable dispersion of clay nanorods on the matrix using various disaggregation techniques, which represents the key to developing functional nanocomposites.⁸²

5. Toxicity of clay minerals

Till date, studies elucidating the toxicological effects of clays at physiological concentrations are not conclusive. Importantly, the oral administration of MMT in rats at high doses (1000 mg/kg) did not lead to accumulation in any organ,⁸³ and cell viability and proliferation remained close to 100% for any concentration of MMT tested in ovarian Hamster cells.⁸⁴ However, low concentrations of MMT (5 µg/mL) in human intestinal cells led to an acute response, inhibiting cell proliferation after 24 h of incubation⁸³, and the same effect was observed in the HepG2 hepatic cell line.⁸⁵

According to Mousa and coworkers,⁸⁶ this behavior is closely related to the flocculation of the clay, as well as the high concentration of salts in the culture medium, which contributed to the formation of agglomerates that accumulated around the cells, leading to cellular damage, such as blockage of membrane channels and alteration of cellular metabolism. Thus, it is appears that

the inhibition of cell proliferation is an indirect effect of clay aggregation rather than a cytotoxic effect of clay itself. This aggregation depends on the surface charge, ion exchange capacity, and the size and morphology of the particles. The authors concluded that the *in vitro* and *in vivo* cytotoxicity studies available clearly showed the biocompatibility of these compounds when they remained stable, i.e., without precipitation. According to the literature^{6,16} these clay materials are inert. However, there is a lack of information concerning their biodistribution and clearance, and if this depends on whether the clays are surgically implanted or administered parentally.

The literature review on the toxicological effects of clays and clay minerals by Maisanaba and coworkers⁸⁷ provided conflicting information, wherein *in vitro* assays generally suggest that clays are cytotoxic, whereas *in vivo* experiments in rodents showed no systemic toxicity. However, the authors concluded that toxicity should be assessed on a case-by-case basis, because it depends on the modifiers used, experimental methodology to assess cytotoxicity, concentration range, purity of the sample, type of deposit used, and its geological formation conditions and time of exposition.

6. Conclusions

Clay minerals have technological advantages in medical sciences provided by their structural, morphological, and textural characteristics. Clay minerals also have several advantages in biosensor applications, controlled release systems, and tissue engineering, especially their biocompatibility and biodegradability. Notably, possible adverse effects of clay minerals on human health remain unclear and could be related to the presence of impurities in the sample, exposure time, or limitations of the experimental biological studies. Such inorganic nanoparticles, either lamellar or fibrous, are expected to be used in association with a wide variety of biomolecules in biotechnological applications. Recent research reinforces the promising potential of clay minerals in the development of biomaterials.

References

- Gill I and Ballesteros A. Bioencapsulation within syn000thetic polymers (Part 1): sol-gel encapsulated biologicals. *Trends in Biotechnology* **18**: 7282-296 (2000).
- Gersbach C. Engineered Bioactive Molecules. *Reference Module in Materials Science and Materials Engineering. Comprehensive Biomaterials* **5**:131-145 (2011).
- An N, Zhou C, Zhuang X, Tong D and Yu W. Immobilization of enzymes on clay minerals for biocatalysts and biosensors. *Appl Clay Sci* **114**: 283-296 (2015).
- Ruiz-Hitzky E, Darder M, Alcántara A, Wicklein B and Aranda P. Recent Advances on Fibrous Clay-Based Nanocomposites. *Adv Polym Sci* **267**: 39-86 (2015).
- Tully J, Yendluri R and Lvov Y. Halloysite Clay Nanotubes for Enzyme Immobilization. *Biomacromolecules* **17**: 615-621 (2016).
- Carretero M, Gomes C and Tateo F. Clays and human health, in *Handbook of Clay Science*, ed by Bergaya F, Theng B and Lagaly G. Elsevier Science, Amsterdam, pp. 717-741 (2006).
- Chrzanowski W, Kim S and Neel E. Biomedical Applications of Clay. *Aust J Chem* **66**:1315-1322 (2013).
- Braun S, Bhattacharyya S and Ducheyne P. Encapsulation of Cells (Cellular Delivery) Using Sol-Gel Silica. *Reference Module in Materials Science and Materials Engineering. Comprehensive Biomaterials II* **5**: 175-186 (2017).
- Williams L, Metge D, Eberl D, Harvey R, Turner A and Prapaipong P. What Makes a Natural Clay Antibacterial? *Environ Sci Technol* **45**: 3768-3773. (2011).
- Ghadiri M, Chrzanowski W and Rohanizadeh R. Biomedical applications of cationic clay minerals. *Roy Soc Ch* **5**: 29467-29481 (2015).
- Jayrajsinh S, Pharm G, Agrawal Y and Bakre L. Montmorillonite nanoclay as a multifaceted drug-delivery carrier: A review. *J Drug Delivery Science and Technology* **39**: 200-209 (2017).
- Yu W, Li N, Tong D, Zhou C, Lin C and Xu C. Adsorption of proteins and nucleic acids on clay minerals and their interactions: A review. *Appl Clay Sci* **80-81**:443-452 (2013).
- Ruiz-Hitzky E, Darder M, Fernandes F, Wicklein B, Alcántara A and Aranda P. Fibrous clays based bionanocomposites. *Prog Polym Sci* **38**: 1392-1414 (2013).
- Dawson J and Oreff R. Clay: new opportunities for tissue regeneration and biomaterial design. *Adv Mater* **25**: 4069-4086 (2013).
- Alcántara A, Darder M, Aranda P and Ruiz-Hitzky E. Polysaccharide-fibrous clay bionanocomposites. *Appl Clay Sci* **96**: 2-8 (2014).
- Aguzzi C, Cerezo P, Viseras C and Caramella C. Use of clays as drug delivery systems: possibilities and limitations. *Appl Clay Sci* **36**: 22-36 (2007).
- Sanchis M, Carsi D, Culebras M, Gómez C, Rodríguez S and Torres F. Molecular dynamics of carrageenan composites reinforced with cloisite Na⁺ montmorillonite nanoclay. *Carbohydr Polym* **176**: 117-126 (2017).
- Dastjerdi R, Mahsa S, Kouros K, Mivehi L and Samadikuchaksaraei A. An acid-free water-born quaternized chitosan/montmorillonite loaded into an innovative ultra-fine bead-free water-born nanocomposite nanofibrous scaffold; in vitro and in vivo approaches. *Biomedical Materials* **12**: 045014 (2017).
- Onnainty R, Onida B, Páez P, Longhi M, Barresi A and Granero G. Targeted chitosan-based bionanocomposites for controlled oral mucosal delivery of chlorhexidine. *Int J Pharma* **509**: 408-418 (2016).
- Moghadas B, Dashtimoghadam E, Mirzadeh H, Farzad S and Mohammad M et al. Novel chitosan-based nanobiohybrid membranes for wound dressing applications. *Rsc Advances* **6**: 7701-7711 (2016).
- Hamidabadi H, Rezvani Z, Bojnordi M, Shirinzadeh H, Seifalian A and Joghataei M. Chitosan-Intercalated Montmorillonite/Poly(vinyl alcohol) Nanofibers as a Platform to Guide Neuronlike Differentiation of Human Dental Pulp Stem Cells. *ACS Appl Mater Inter* **9**: 11392-11404 (2017).
- Bera H, Ippagunta S, Sanoj K and Vangala P. Core-shell alginate-ghatti gum modified montmorillonite composite matrices for stomach-specific flurbiprofen delivery. *Mater Sci Eng C* **76**: 715-726 (2017).
- Alavijeh M, Sarvi M and Afarani Z. Properties of adsorption of vitamin B12 on nanoclay as a versatile carrier. *Food Chemistry* **219**: 207-214 (2017).
- Saha N, Sarkar G, Roy I, Rana D, Bhattacharyya A, Adhikari A et al. Studies on methylcellulose/pectin/montmorillonite

- nanocomposite films and their application possibilities. *Carbohydr Polym* **20**: 1218-1227 (2016).
25. Dening T, Joyce P, Rao S, Thomas N and Prestidge C. Nanostructured Montmorillonite Clay for Controlling the Lipase-Mediated Digestion of Medium Chain Triglycerides. *Acs Appl Mater Inter* **8**: 32732-32742 (2016).
 26. Costa M, Ferreira I and Cruz M. New polyelectrolyte complex from pectin/chitosan and montmorillonite clay. *Carbohydr Polym* **146**: 123-130 (2016).
 27. Wilpiszewska K, Antosik A and Spychaj T. Novel hydrophilic carboxymethyl starch/montmorillonite nanocomposite films. *Carbohydr Polym* **128**: 82-89 (2015).
 28. Rangel-Rivera P, Rangel-Porrás G, Pfeiffer-Perea H and Lima-Muñoz E. Thermoanalytical study of acid-treated clay containing amino acid immobilized on its surface. *J Therm Anal Calorim* **115**: 359-1369 (2014).
 29. Kędzierski M, Janiszewska J and Moszumańska I. Dendrimeric peptide-montmorillonite intercalation compound. *Polimery* **61**: 677-682 (2016).
 30. Santana H, Paesano Jr A, Costa A, Mauro E, Souza I and Ivashita F. et al. Cysteine, thiourea and thiocyanate interactions with clays: FT-IR, Mössbauer and EPR spectroscopy and X-ray diffractometry studies. *Amino Acids* **38**: 1089-1099 (2010).
 31. Beall G, Sowersby D, Roberts R, Robson M and Lewis L. Analysis of oligonucleotide DNA Binding and sedimentation properties of montmorillonite clay using ultraviolet light spectroscopy. *Biomacromolecules* **10**: 105-112 (2009).
 32. Sangwan P, Way C and Wu D-Y. New Insight into Biodegradation of Polylactide (PLA)/Clay Nanocomposites Using Molecular Ecological Techniques. *Macromol Biosci* **9**: 677-686 (2009).
 33. Ralla K, Sohling U, Riechers D, Kasper C, Ruf F and Scheper T. Adsorption and separation of proteins by a smectitic clay mineral. *Bioproc Biosyst Eng* **33**: 847-861 (2010).
 34. Charradi K, Gondran C, Amara A, Prevot V and Mousty, C. H₂O₂ determination at iron-rich clay modified electrodes. *Electrochim Acta* **54**: 4237-4244 (2009).
 35. El Adraa K, Timon V, Lambert J-F, Al-Rabaa A-R, Jaber F, Jaber M. et al. Adsorption of L-DOPA Intercalated in Hydrated Na-Saponite Clay: A Combined Experimental and Theoretical Study. *J Phys Chem C* **116**: 26414-26421 (2012).
 36. Postnova I, Sarin S, Silant'ev V, Ha C and Shchipunov Y. Chitosan bionanocomposites prepared in the self-organized regime. *Pure Appl Chem* **87**: 793-804 (2015).
 37. Benetoli L, Souza C, Silva K, Souza Jr I, Santana H, Paesano Jr A et al. Amino acid interaction with and adsorption on clays: FT-IR and Mössbauer spectroscopy and X-ray diffractometry investigations. *Orig Life Evol Biosph* **37**: 479-93 (2007).
 38. Barua S, Dutta N, Karmakar S, Chattopadhyay P, Aidew L and Buragohain A. et al. Biocompatible high performance hyperbranched epoxy/clay nanocomposite as an implantable material. *Biomed Mater* **9**: 025006 (2014).
 39. Bonifacio M, Gentile P, Ferreira A, Cometa S, De Giglio E. Insight into halloysite nanotubes-loaded gellan gum hydrogels for soft tissue engineering applications. *Carbohydr Polym* **163**: 280-291 (2017).
 40. Bertolino V, Cavallaro G, Lazzara G, Milioto S and Parisi F. Biopolymer-Targeted Adsorption onto Halloysite Nanotubes in Aqueous Media. *Langmuir* **33**: 3317-3323 (2017).
 41. Huang B, Liu M and Zhou C. Cellulose-halloysite nanotube composite hydrogels for curcumin delivery. *Cellulose* **24**: 2861-2875 (2017).
 42. Wahab I and Abd Razak S. Bionanocomposite Film of Kappa-Carrageenan/Nanotube Clay: Growth of Hydroxyl Apatite and Model Drug Release. *Dig J Nanomater Bios* **11**: 963-972 (2016).
 43. Darder M, Matos C, Aranda P, Gouveia R and Ruiz-Hitzky, E. Bionanocomposite foams based on the assembly of starch and alginate with sepiolite fibrous clay. *Carbohydr Polym* **157**: 1933-1939 (2017).
 44. Gur E, Altinisik A and Yurdakoc K. Preparation and characterization of chitosan/sepiolite bionanocomposites for tetracycline release. *Polym Composite* **38**: 1810-1818 (2017).
 45. Sárossy Z, Blomfeldt J, Hedenqvist M, Koch C, Ray S and Plackett D. Composite Films of Arabinoxylan and Fibrous Sepiolite: Morphological, Mechanical, and Barrier Properties. *ACS Appl Mater Inter* **4**: 3378-3386 (2012).
 46. Ruiz-Hitzky E, Darder M, Aranda P, Martín del Burgo M, del Real G. Virus-bionanocomposite materials: applications for flu vaccines. *Adv Mater* **21**: 4167-4171 (2009).
 47. Wang Z, Zhao Y, Luo Y, Wang S, Shen M, Tomás H et al. Attapulgit-doped electrospun poly(lactic-co-glycolic acid) nanofibers enable enhanced osteogenic differentiation of human mesenchymal stem cells. *RSC Advances* **5**: 2383-2391 (2015).
 48. Alcántara A, Darder M, Aranda P, Ruiz-Hitzky E. Zein-fibrous clays biohybrid materials. *European J Inorganic Chem* **32**: 5216-5224 (2012).
 49. Yang H, Wang W, Wang A. A pH-sensitive biopolymer-based superabsorbent nanocomposite from sodium alginate and attapulgite: synthesis, characterization, and swelling behaviors. *J Disper Sci Technol* **33**: 1154-1162 (2012).
 50. Wang Q, Zhang J and Wang A. Preparation and characterization of a novel pH-sensitive chitosan-g-poly (acrylic acid)/attapulgite/sodium alginate composite hydrogel bead for controlled release of diclofenac sodium. *Carbohydr Polym* **78**: 731-737 (2009).
 51. Chen G-J, Yen M-C, Wang J-M, Lin J-J and Chiu H-C. Layered Inorganic/Enzyme Nanohybrids with Selectivity and Structural Stability upon Interacting with Biomolecules. *Bioconjugate Chem* **19**: 138-144 (2008).
 52. Mousty C. Sensors and biosensors based on clay-modified electrodes — new trends. *Appl Clay Sci* **27**: 159-177 (2004).
 53. Ghosh P and Bard A. Clay-modified electrodes. *J Am Chem Soc* **105**: 5691-5693 (1983).
 54. Navrátilová Z and Kula P. Clay modified electrodes: present applications and prospects. *Electroanalysis* **15**: 837-846 (2003).
 55. Mousty C. Biosensing applications of clay-modified electrodes: a review. *Anal Bioanal Chem* **396**: 315-325 (2010).
 56. Mousty C and Prévot V. Hybrid and biohybrid layered double hydroxides for electrochemical analysis. *Anal Bioanal Chem* **405**: 3513-3523 (2013).
 57. Zhao L, Zhou C, Wang J, Tong D, Yu W and Wang H. Recent advances in clay mineral-containing nanocomposite hydrogels. *Soft Matter* **11**: 9229-9246 (2015).
 58. Mousty C and Walcarius A. Electrochemically assisted deposition by local pH tuning: a versatile tool to generate ordered mesoporous silica thin films and layered double hydroxide materials. *J Solid State Electr* **19**: 1905-1931 (2015).
 59. Ramachandran R, Chen S-M, Kumar G, Gajendran P, Xavier A and Devi N. High Electroactive Electrode Catalysts and Highly sensitive Electro analytical Techniques for Hydrogen Peroxide detection. *Int J Electr Sci* **11**: 1247-1270 (2016).
 60. Saifullah B and Hussein M. Inorganic nanolayers: structure, preparation, and biomedical applications. *Int J Nanomed* **10**: 5609-5633 (2015).
 61. Unal B, Yalcinkaya E, Gumustas S, Sonmez B, Ozkan M, Balcan M et al. Polyglycolide-montmorillonite as a novel nanocomposite platform for biosensing applications. *New J Chem* **41**: 9371-9379 (2017).
 62. Yadav D, Gupta R, Ganesan V and Sonkar P. Individual and simultaneous voltammetric determination of ascorbic acid, uric acid and folic acid by using a glassy carbon electrode modified with gold nanoparticles linked to bentonite via cysteine groups. *Microchim Acta* **184**: 1951-1957 (2017).
 63. Liu M, He R, Yang J, Zhao H and Zhou C. Stripe-like Clay Nanotubes Patterns in Glass Capillary Tubes for Capture of Tumor Cells. *ACS Appl. Mater. Inter* **8**: 7709-7719 (2016).
 64. Fernandes F and Ruiz-Hitzky E. Assembling nanotubes and nanofibres: Cooperativeness in sepiolite-carbon nanotube materials. *Carbon* **72**: 296-303 (2014).
 65. Demir F, Demir B, Yalcinkaya E, Cevik S, Demirkol D, Anik U. et al. Amino acid intercalated montmorillonite: electrochemical biosensing applications. *RSC Advances* **4**: 50107-50113 (2014).
 66. Sonmez B, Sayin S, Yalcinkaya E, Selecı D, Yildiz H, Demirkol D. et al. Calixarene modified montmorillonite: a novel design for biosensing applications. *RSC Advances* **4**: 62895-62902 (2014).
 67. Songurtekin D, Yalcinkaya E, Ag D, Selecı M, Demirkol D and Timur S. Histidine modified montmorillonite: Laccase immobilization and applications to flow injection analysis of phenols. *Appl Clay Sci* **86**: 64-69 (2013).
 68. Luo S, Chen Y, Zhuo M, Yao C, Xi H, Kong Y. et al. Palygorskite-poly(o-phenylenediamine) nanocomposite: An enhance electrochemical platform for glucose biosensing. *Appl Clay Sci* **86**: 59-63 (2013).
 69. Chen H, Zhang Z, Cai D, Zhang S, Zhang B and Tang J. Direct electrochemistry and electrocatalytic behavior of horseradish peroxidase on attapulgite clay modified electrode. *Analytical Sci* **27**: 613-616 (2011).
 70. Chen J and Jin Y. Sensitive lactate determination based on acclimated mixed bacteria and palygorskite co-modified oxygen electrode. *Bioelectrochemistry* **80**: 151-154 (2011).
 71. Wu P, Cai Z, Chen J, Zhang H and Cai C. Electrochemical measurement of the flux of hydrogen peroxide releasing from RAW 264.7 macrophage cells based on enzyme-attapulgite clay nanohybrid. *Biosens Bioelectron* **26**: 4012-4017 (2011).
 72. Chen, J and Jin, Y. Sensitive phenol determination based on co-modifying tyrosinase and palygorskite on glassy carbon Electrode. *Microchim Acta* **169**: 249-254 (2010).
 73. Xu J, Han W, Yin Q, Song J and Zhong H. Direct Electron Transfer of Glucose Oxidase and Glucose Biosensor Based on Nano-structural Attapulgite Clay Matrix. *Chinese J Chem* **27**: 2197-2202 (2009).
 74. Xu J, Li W, Yin Q, Zhong H, Zhu Y and Jin L. Direct electron transfer and bioelectrocatalysis of hemoglobin on non-structural attapulgite clay-modified glassy carbon Electrode. *J Colloid Interf Sci* **315**: 170-176 (2007).
 75. Xu J, Li W, Yin Q and Zhu Y. Direct electrochemistry of Cytochrome c on natural nano-attapulgite clay modified electrode and its electrocatalytic reduction for H₂O₂. *Electrochim Acta* **52**: 3601-3606 (2007).
 76. Kong Y, Chen X, Wang W and Chen Z. A novel palygorskite-modified carbon paste amperometric sensor for catechol determination. *Anal Chim Acta* **688**: 203-207 (2011).
 77. Bramhill J, Ross S and Ross G. Bioactive Nanocomposites for Tissue Repair and Regeneration: A Review. *Int J Environ Res Public Health* **14**: 66 (2017).
 78. Mohd S, Abdullah M and Amin K. Gellan gum/clay hydrogels for tissue engineering application: Mechanical, thermal behavior, cell viability and antibacterial properties. *J Bioact Compat Pol* **31**: 1-19 (2016).
 79. He L, Ding K, Wu H, Wang N, Williams C, Yan Q et al. Evaluation of Cytotoxicity of natural nano-Attapulgite and its enhancement of Vero Cell Productivity. *Dig J Nanomater Bios* **8**: 551-560 (2013).
 80. Zafar R, Zia K, Tabasum S, Jabeen F, Noreen A and Zuber M. Polysaccharide based bionanocomposites, properties and applications: A review. *Int J Biol Macromol* **92**: 1012-1024 (2016).
 81. Cervini-Silva J, Nieto-Camacho A, Ramírez-Apan M, Gómez-Vidales V, Palacios E, Montoya A et al. Anti-inflammatory, anti-bacterial, and cytotoxic activity of fibrous clays. *Colloid Surfac B* **129**: 1-6 (2015).
 82. Wang W and Wang A. Recent progress in dispersion of palygorskite crystal bundles for nanocomposites. *Appl Clay Sci* **119**: 18-30 (2016).
 83. Baek M, Lee J and Choi S. Toxicological effects of a cationic clay, montmorillonite in vitro and in vivo. *Mol Cell Toxicol* **8**: 95-101 (2012).
 84. Li P, Wei J, Chiu Y, Su H, Peng F, Lin J. Evaluation on cytotoxicity and genotoxicity of the exfoliated silicate nanoclay. *ACS Appl Mater Inter* **2**: 1608-1613 (2010).
 85. Lordan S, Kennedy J and Higginbotham C. Cytotoxic effects induced by unmodified and organically modified nanoclays in the human hepatic HepG2 cell line. *J Appl Toxicol* **31**: 27-35 (2011).
 86. Mousa M, Evans N, Oreffo R and Dawson J. Clay nanoparticles for regenerative medicine and biomaterial design: a review of clay bioactivity. *Biomaterials* **159**: 204-214 (2018).
 87. Maisanaba S, Pichardo S, Puerto M, Gutiérrez-Praena D, Cameán A and Jos A. Toxicological evaluation of clay minerals and derived nanocomposites: a review. *Environ Res* **138**: 233-254 (2015).